Abstract

Rates of skin cancer continue to increase despite the improved use of traditional sunscreens to minimize damage from ultraviolet radiation. The public perception of tanned skin as being healthy and desirable, combined with the rising demand for treatments to repair irregular skin pigmentation and the desire to increase or decrease constitutive skin pigmentation, arouses great interest pharmaceutically as well as cosmeceutically. This review discusses the intrinsic biochemistry of pigmentation, details mechanisms that lead to increased or decreased skin pigmentation, and summarizes established and potential hyper- and hypo-pigmenting agents and their modes of action.

Keywords: skin, pigmentation, ultraviolet, hypopigmentation, hyperpigmentation

Introduction

Despite the increased use of traditional sunscreens, the incidence of malignant melanoma has tripled over the past 40 years. In addition to the progressive thinning of the ozone layer, increased recreational sun exposure of the population and the cosmetically motivated desire to tan are held at least in part responsible. The widely accepted view that sunscreens protect against melanoma has been challenged by epidemiological and experimental studies that revealed no or even an inverse association between the use of sunscreens and melanoma development [1,2]. In addition, there is a rising demand for remedies to treat irregular hyperpigmentation or, especially in Asian cultures, to obtain a general whitening of the skin. In light of these facts, the modification of skin pigmentation, whether by hyper-or
hypo-pigmenting agents, arouses great interest pharmaceutically as well as cosmeceutically. This review discusses the intrinsic biochemistry of pigmentation, details mechanisms that lead to increased or decreased pigmentation and summarizes established and potential hyper- and hypopigmenting agents and their modes of action.

Biochemistry of melanogenesis

Skin color is determined by a mixture of four biochromes: oxyhemoglobin (red), reduced hemoglobin (blue), carotenoids (yellow) and, most importantly, the amounts and types of melanins (brown) produced and their distribution in the skin. Although the number of melanocytes (specialized cells in the epidermis that produce the pigment) does not differ between subjects from different racial/ethnic backgrounds [3], there are intra-individual differences in melanocyte density at different sites of the body (e.g. the palms and soles contain only 10–20% the density of melanocytes found elsewhere) [4]. Melanocytes account for only 1% of epidermal cells and occur at an approximate ratio of 1:10 among keratinocytes in the basal skin layer. Via their elongated dendrites, melanocytes transport their ovoid membrane-bound organelles (termed melanosomes), in which melanin is synthesized and stored, to neighboring keratinocytes [5], where melanosomes form a critical barrier as supranuclear “caps” to shield DNA from ultraviolet radiation (UVR) [6]. Proliferating keratinocytes in the suprabasal epidermal layers gradually ascend towards the skin surface along with their ingested melanin to contribute to photoprotection.

The regulation of pigmentation is a complex process and there are currently more than 125 genes known to be directly or indirectly involved [7]. Many genes (>25) regulate melanosomal biogenesis or function, which requires a number of specific enzymatic and structural proteins for effective melanin production [8,9].

Three different kinds of melanin are produced by melanocytes: the lighter red/yellow, alkali soluble sulfur-containing pheomelanin (which is predominant in the red hair/freckles phenotype) and two types of eumelanin, dark brown/black insoluble pigments found in dark skin and black hair. Human skin normally contains a mixture of all 3 types of melanin, but the ratio varies greatly and determines the color of the skin [10].

Within melanosomes, at least 3 enzymes are indispensable for the synthesis of different types of melanin (Figure 1). Tyrosinase (TYR) catalyzes the initial rate-limiting step in melanogenesis, the hydroxylation of tyrosine to β-3,4-dihydroxyphenylalanine (DOPA) and the subsequent oxidation of DOPA to DOPAquinone. In the presence of cysteine, DOPAquinone is converted stochiometrically into 3- or 5-cysteinyldOPA which oxidizes and polymerizes to pheomelansins [11,12]. After depletion of cysteine, DOPAquinone spontaneously cyclizes to form DOPAchrome. Dark brown/black 5,6-dihydroxyindole (DHI) melansins are generated after DOPAchrome spontaneously loses its carboxylic acid group and oxidizes and polymerizes. However, in the presence of the enzyme DOPAchrome tautomerase (DCT), the carboxylic acid group is retained, and DOPAchrome is converted into DHI-2-carboxylic acid (DHICA), which finally results in the formation of DHICA-melanin, a moderately soluble lighter brown product of intermediate size [13].
Tyrosinase-related protein 1 (TYRP1) is a critical enzyme for the correct trafficking of tyrosinase to melanosomes [14], and DCT also seems to play an important role in detoxification processes within melanosomes [15]. Mutations in these enzymes lead to dramatic changes in the quantity and quality of synthesized melanin. Tyrosinase is responsible for the critical rate-limiting steps of melanogenesis; mutations that affect TYR function result in albinism. Mutations in proteins that are involved in sorting/trafficking of proteins to melanosomes or in melanosome structure result in inherited hypopigmentary disorders [16]. Proteins regulating the uptake of substrates needed for melanin production, for example tyrosine and cysteine transporters, as well as those regulating the pH within melanosomes, for example proton pumps, are also critical in regulating melanin production.

**Figure 1**

Schematic of the melanin synthetic pathway and enzymes involved that occur specifically in melanosomes

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**Regulation of constitutive pigmentation**

The determination of pigmentary phenotype is genetically complex and at a physiological level is quite complicated. Genes determining a number of rare Mendelian disorders of pigmentation, such as albinism, have been identified, but one gene, the melanocortin-1 receptor (MC1R), has been found to explain much of the variation of skin color in the normal population. The MC1R is a G protein coupled receptor expressed on the surface of melanocytes that regulates the quality and quantity of melanin production [17,18]. MC1R is regulated by agonists (α-melanocyte stimulating hormone (αMSH) and adrenocorticotrophic hormone (ACTH)) as well as by an antagonist (agouti-signaling protein (ASP)). Binding of an agonist to MC1R triggers the melanogenic cascade that starts with the activation of adenylate cyclase that results in the stimulation of cyclic adenosine monophosphate (cAMP) synthesis (Figure 2), activation of the cAMP-dependent protein kinase A (PKA) and then a series of reactions (many of them are yet unknown) that finally result in the increased synthesis of eumelanin. If the MC1R signal is blocked by ASP, the eumelanin synthesis switches to pheomelanin production. The switch to synthesize eu-versus pheomelanin is controlled by MC1R, but the mechanism(s) remain obscure. The high polymorphism of MC1R accounts for the diversity of constitutive pigmentation from extremely fair to extremely dark skin while defective MC1R variants that lead to decreased function of the receptor that have been associated with red hair and fair skin [19] and high sun sensitivity [19,20] as well as an increased risk of melanoma [21]. Beyond MC1R control, various environmental factors such as UVR and factors secreted by surrounding keratinocytes and fibroblasts in the skin can influence melanocyte proliferation and differentiation significantly. DKK1, a factor secreted by dermal fibroblasts in the palms/soles is an inhibitor of the Wnt/β-catenin pathway and therefore able to suppress melanocyte...
growth and function dramatically [4,22]. Not only does DKK1 inhibit the expression of transcription factors (e.g. MITF) and melanogenic proteins, but it also impairs the ability of keratinocytes for melanin uptake.

Figure 2

There are other factors that influence melanin production, such as P and MATP proteins; both are melanocyte-specific 12 membrane-transporters that are key factors in the sorting/trafficking of tyrosinase to melanosomes. The exact mechanisms of P and MATP remain to be elucidated, but it has been hypothesized that they act as transporters/pumps that control intracellular ion transport and in this way regulate tyrosinase sorting and melanin synthesis. As shown by population studies, polymorphisms of the P [23] and MATP [24] genes (in combination with polymorphisms of the MC1R gene) make major contributions to determine the normal range of pigmentation in hair, skin and eyes. Recently, an important role of SLC24A5, a member of a family of potassium-dependent sodium-calcium exchangers, in the regulation of pigmentation was reported. A close association was found between a polymorphism in SLC24A5 (Ala111Thr) and cutaneous melanin content in an intermixed group [25]. On the basis of the HapMap database (www.hapmap.org), the fixed Ala111Thr polymorphism of SLC24A5 was found in Europeans (with lighter skin color) while the ancestral allel (Ala111) was conserved in African-Americans and in African-Caribbeans (with darker skin). Dysfunctions of TYR and/or other proteins of the melanogenic pathway lead to the disruption of skin, hair and eye pigmentation, a condition termed albinism [26]. So far, 5 types of albinism have been described that are associated with 5 distinct pigment-related loci. Mutations in those genes result in direct or indirect impairment of TYR activity. Proteasomal degradation of TYR is characteristic for oculocutaneous albinism (OCA) type 1 (TYR) and OCA3 (TYRP1), while OCA2 (P) and OCA4 (MATP) lead to disrupted sorting of functional TYR to melanosomes. Ocular albinism type 1 (OA1) causes a disruption of melanin synthesis, primarily in the eye, by an as yet unknown mechanism [27].
In sum, constitutive pigmentation depends on: (a) melanocyte number, (b) the expression and function of melanosomal enzymes and structural proteins, (c) the amount of eu- and pheomelanin synthesized, (d) the transport of melanosomes to dendrites, (e) their transfer to keratinocytes and, (f) last but not least, the distribution of melanin in suprabasal skin layers.

Regulation of facultative pigmentation

An increase of skin pigmentation above the basal constitutive level induced by physiological factors (for example, UVR) is called facultative pigmentation, or more commonly ‘tanning’ [28]. The relationship between MC1R and UVR is complex and is regulated at many levels [29]. Microphthalmia-associated transcription factor (MITF - the master regulator of pigmentation) is the most important factor involved in the MC1R regulation of melanocyte function. Levels of MITF are quickly increased by UVR which then stimulates the expression of downstream melanogenic proteins, including Pmel17, MART1, TYR, TYRP1 and DCT [30,31]. The sum of those factors eventually leads to significant increases in melanin content after several weeks.

UVR also leads to increased melanocyte density, melanin synthesis and melanocyte dendricity, which play important roles in the transfer of melanin to keratinocytes. UV also enhances levels of proteinase-activated receptor 2 (PAR2) which stimulates melanosome uptake and distribution by epidermal keratinocytes [32]. Melanocytes, keratinocytes and fibroblasts react to UVR by releasing a wide variety of melanogenic factors. Following exposure to UVR, melanocytes as well as keratinocytes express higher amounts of proopiomelanocortin (POMC), the precursor of the melanocortins αMSH and ACTH, which stimulates MC1R function and consequently boosts melanocyte responses to those melanocortins. Keratinocytes also respond to UV with increased expression of other factors, including endothelin-1 (ET-1) which stimulates melanocyte function in a paracrine manner [33], interleukin-1 (IL-1) which enhances autocrine secretion of ACTH, αMSH, ET-1 and basic fibroblast growth factor (bFGF), stem cell factor (SCF) which is involved in the regulation of proliferation and melanogenesis of melanocytes, and nerve growth factor (NGF) which is involved in regulating melanogenesis and/or dendritogenesis of melanocytes [34]. UV-induced activation of p53 in keratinocytes leads to increased expression of the POMC gene which in turn results in increased levels of αMSH and stimulation of MC1R function in melanocytes [35]. Fibroblast activation by UVR results in the secretion of growth factors, including hepatocyte growth factor (HGF), bFGF and SCF all of which stimulate pigmentation via their receptors on melanocytes [36]. Hormones also represent important factors that regulate skin pigmentation. It was hypothesized that vitamin D3, which is synthesized in the skin upon UV exposure, might play a role in the melanogenic effects of UV, as human melanocytes also express vitamin D3 receptors, but so far, the underlying mechanism(s) remains to be elucidated. For example, topical application of 1,25-(OH)2-vitamin D3 leads to an increase in the number of melanocytes and augments the melanogenic effect of UV on mouse skin, while melanocytes in vitro respond to 1,25-(OH)2-vitamin D3 by decreasing TYR activity [37]. Sex steroid hormones also play an important role in skin pigmentation. In response to β-estradiol, melanocytes increase TYR activity in a dose-dependent manner although there is no correlation of this response with constitutive pigmentation [38,39].
Although the mechanisms involved in pigmentation are similar in skin of different racial/ethnic groups, melanin transfer from the basal level of the epidermis upwards is more effective in dark skin than in fair skin resulting in a better protection of the lower epidermis in dark skin [40,41].

**Induction of hyperpigmentation – specific mechanisms**

In the face of the complex mechanisms that regulate human skin pigmentation, disorders that result from exogenous or endogenous influences are not uncommon. As a vast variety of factors exists that can lead to hyper- or hypo-pigmentation, this review will focus on the more common and well-investigated stresses that can lead to acquired pigmentation disorders. Increased levels of melanin in the epidermis result in a state known as hypermelanosis. Two types of changes exist: a) increased numbers of melanocytes in the epidermis followed by increased production of melanin, which is called melanocytic hypermelanosis (e.g. lentigo), and b) no increase of melanocyte number but increased melanin production only, termed melanotic hypermelanosis (e.g. melasma). Hypermelanosis of both types can be the result of genetic, hormonal (increase in circulating pituitary melanotropic hormones) and environmental (UVR) factors.

UV is one of the most powerful agents that induces hyperpigmentation of the skin. Lentigines solares (LS) (also termed age spots, sun spots and actinic lentigines) are circumscribed, pigmented macules, usually brown in color, that range in size from a few millimeters to a few centimeters in diameter and may coalesce into even more extended lesions [42]. They typically appear on sun-exposed areas of the skin such as the neck, face and forearms [43] and increase in number with age, affecting more than 90% of the Caucasian population older than 50 years [44]. LS result from increased amounts of melanin in the basal and suprabasal layers of the epidermis. The mechanisms underlying this type of hyperpigmentation process have recently been elucidated by Imokawa and coworkers. Besides a two-fold increase of TYR-positive melanocytes in lesional skin compared to perilesional skin [45], they demonstrated the existence of a molecular network in which increased expression of the ET-1/ET(B)R cascade and higher expression of SCF in lesional skin as well as cross-talk between these two signaling pathways following UV exposure play an important role in the mechanisms underlying LS [46,47].

A wide variety of drugs and substances has been reported to induce hyperpigmentation, including antibiotics (mainly tetracyclines), chemotherapeutics, heavy metals and antiepileptic medications. Diffuse muddy brown discolorations in sun-exposed areas of the skin (type III reaction) induced by minocyclin, a tetracycline-derivative, are well documented side-effects presumably resulting from increased melanin production by minocyclin-stimulated melanocytes that can lead, among other things, to deposits of melanin or minocyclin/melanin-complexes at the epidermal basal membrane and in the dermis [48]. Chemotherapeutics such as bleomycin, daunorubicin, doxorubicin, cyclophosphamide and 5-floururacil are able to cause hyperpigmentation, supposedly by stimulation of melanogenesis via direct toxic effects on melanocytes, although the underlying mechanisms are unknown. Based on the observation that fragments of nucleic acids can stimulate melanin synthesis [49], chemotherapy-induced damage to DNA in skin cells could induce signals that promote melanogenesis [50]. Heavy metals such as gold, silver, arsenic or bismuth can enhance melanin synthesis [51]. It is believed that such metals
complex with sulfhydryl compounds in the skin that normally block TYR activity and thus resulting in the stimulation of melanogenesis. Antiepileptic drugs (e.g. hydantoins) are also known to increase pigmentation and might do so by a direct stimulatory action on melanocytes [52]. Tricyclic depressants (desipramine and imipramine) are associated with slate-gray pigmentation in sun-exposed areas caused both by increased melanin in the dermis as well as by electron-dense inclusions within dermal cells [53,54].

Melasma is a hyperpigmentation disorder that presents with arcuate or polycyclic hyperpigmented lesions in sun-exposed areas and occurs most commonly in women in the central facial area [55]. So far, the cause of melasma is not known but a large number of factors exist that can contribute to its development or aggravation (e.g. pregnancy and oral contraceptive/hormone replacement therapy, UV exposure, genetic influences, and cosmetics). Among those factors, UVR is regarded as the most important cause [56]. It was shown that lesional skin of melasma has higher amounts of melanin in the epidermis and dermis but no increase in melanocyte number, although the melanocytes were larger and more dendritic and produced higher amounts of eumelanin [57]. Elevated levels of estrogen and progesterone are associated with melasma [58] and several studies have shown a stimulating effect of estrogens on tyrosinase activity in cultured melanocytes [38,39].

In almost all skin disorders with an inflammatory component (e.g. acne, eczema, and psoriasis) there is a risk of postinflammatory hyperpigmentation (PIH) due to excessive cytokine secretion [59]. As with melasma, PIH occurs more often in patients with darker skin, although there is no gender predominance. PIH lesions, which may involve the epidermis and/or the dermis, are characteristically limited to sites of preceding inflammation and have hazy, feathered borders. In PIH, the number of melanocytes is typically normal, but the production of melanin is markedly increased. Often, melanin is abnormally transported into the dermis (termed pigmentary incontinence) where it accumulates within melanophages. During inflammatory processes and also after UVR, keratinocytes produce prostaglandins (PG), a group of inflammatory mediators, that are metabolites of arachidonic acid. PGE1 and PGE2 increase melanogenesis strongly while PGA1 and PGD2 represent strong inhibitors [60]. Leukotrienes such as LTC4 and LTD4, are metabolites of the lipoxygenase pathway and are able to induce the proliferation of melanocytes in vitro [61]. Histamine, another inflammatory agent, may activate melanogenesis via proteinase A activation [62].

**Induction of hypopigmentation – specific mechanisms**

Decreased levels of melanin in the epidermis is called hypomelanosis and results mainly from two different types of changes: a) decreased numbers or absence of melanocytes in the epidermis resulting in little or no melanin production (melanocytopenic hypomelanosis, e.g. vitiligo), and b) no decrease in the number of melanocytes but decreased levels of melanin production (melanopenic hypomelanosis). Hypomelanosis can also result from genetic (as in albinism), from autoimmune, toxic or inflammatory processes. During the senescence process, the density of melanocytes in the skin decreases physiologically ~10% per decade [63], but loss of pigmentation can occur at all ages after exposure to melanotoxic agents. Contact with certain chemicals can lead to cutaneous depigmentation [64–66] and
a recent description of all agents that can induce depigmentation of the skin is available [67]. The majority of these agents are derivatives of phenol and catechols [68]. Generally, these agents induce chemical leucoderma (depigmentation at the contact sites), but in subjects with a genetic predisposition and chronic exposure, the initial depigmentation can extend and lead to progressive generalized vitiligo. Large differences in the appearance of toxic effects after exposure have been observed, and it seems that there is a genetic control of responses to melanotoxic agents [68].

Hypomelanosis can occur postinflammation (e.g. after resolving of psoriatic plaques or lesions of atopic dermatitis), possibly resulting from an increased keratinocyte turnover that interferes with melanosomal transfer as well as the activation of inhibitory cytokines. Decreased melanin content can be also related to infections (e.g. lepra and syphilis), although the mechanism(s) involved is currently unknown and it has been suggested to be postinflammatory [69]. Although there are some indications that properties of infectious agents might be responsible for the depigmentation: mycobacterium leprae contains an enzyme similar to TYR that potentially converts DOPA to a quinone, so that DOPA is unavailable for melanin production [70].

There are many different possible mechanisms for hypomelanocytosis, including the aforementioned melanotoxic effects of chemicals, or trauma. In vitiligo, there are several hypotheses to explain the loss of melanocytes, including the concept that vitiligo is an autoimmune response by anti-melanocyte antibodies [71] or is mediated by T-cells [72], or is an autotoxic response by melanin precursors [73], as well as being a genetic disease [74].

Modifying skin pigmentation

1. Stimulation of melanogenesis

Compared to fair skin, darkly pigmented skin has a up to 70-fold higher protection against skin cancer [75]. For decades, scientists have tried to enhance skin pigmentation without UV exposure to confer the protective properties of a tan without the associated DNA damage that is caused by UVR. Space doesn’t allow a full elaboration of all agents tested (many of them have been used in vitro only), so we will focus on some with different modes of actions that present interesting approaches to stimulate pigmentation in human skin. Interested readers are referred to a more detailed review that examined enhancers of skin pigmentation [76]. One possible approach lies in triggering MC1R function. In the 1960s, Lerner and McGuire discovered that injections with αMSH, one of the MC1R agonists (see Figure 2), increased human skin pigmentation [77]. More recently, several studies examined the effects of [Nle4-d-Phe7]-αMSH, a synthetic superpotent analogue of αMSH, on human skin in situ and reported increased pigmentation after a series of 10 injections [78–81]. In a larger study, injections of 65 subjects with a slow-release formulation of the same αMSH analogue over 3 months lead to an average 41% increase of melanin in subjects with high sun-sensitivity compared to a 12% increase in subjects with low sun-sensitivity, and there was no significant difference in pigmentation between sun-exposed and non-sun-exposed areas [82]. As the necessity to inject the drug on a regular basis to maintain a tan is a major drawback, there are currently attempts to develop αMSH analogs that are small enough to reach their target when administered topically. Abdel-Malek and colleagues developed potent tetrapeptide αMSH
analogs (n-Pentadecanoyl- and 4-Phenylbutyryl-His-D-Phe-Arg-Trp-NH2) that are able to stimulate melanogenesis and enhance DNA repair after UVR of melanocytes in vitro [83]. However, αMSH analogs may not work in subjects with red hair who have an impaired MC1R function. A different approach was recently demonstrated by D’Orazio and colleagues in an animal model [84]. They were able to induce artificial tanning in red/blonde-haired mice that have an inactivating mutation of MC1R by treating the animals topically with forskolin. Forskolin, a cell permeable diterpenoid, is a natural product (root extract of Plectranthus barbatus, also known as Coleus forskohlii) that bypasses MC1R function by activating adenylate cyclase [85] and thus increasing cAMP levels. This chemically induced tan was able to protect UV-irradiated mice against sunburn, DNA damage and subsequent carcinogenesis. As virtually all cells contain adenylate cyclase, attempts are now being made to develop agents that affect the melanogenic pathway more specifically. UVR not only stimulates αMSH secretion but it also causes DNA damage, and it is widely believed that such damage can itself induce tanning and DNA repair responses [86]. Damage to telomere loops at the ends of chromosomes and overhang exposure is considered to be a DNA damage signal [87]. Oligonucleotides that imitate this telomere overhang (T-oligos) presumably induce protective DNA damage responses. Gilchrest and coworkers found that in human skin explants treated with T-oligos and irradiated with UVB, there was a strikingly reduced amount of DNA damage and a 3–5 fold increase in melanin content compared to untreated UV-irradiated samples [88]. Sunscreens containing T-oligos could be efficient in enhancing skin pigmentation and in protecting against photodamage and skin cancer. Bicyclic monoterpenene (BMT) diols are small molecule compounds found in abundance in plants and food. In cultured cells and in guinea pig skin [89] and to some extent in human skin, BMT diols were able to increase melanin content when combined with α-hydroxy acids or retinoids [76]. BMT diols seem to have a good safety profile and might be interesting agents for further exploration as pigmentation enhancers.

2. Inhibition of pigmentation

The management of skin hyperpigmentation is still a challenging matter for dermatologists, as they are confronted with numerous different therapeutic options which often show unsatisfactory effects. As a further complication, relapses in hyperpigmentary disorders are common. Considering the numerous agents that have been suggested to have a hypopigmenting effect (and many of the available data are for in vitro results only), this review can only be selective in its coverage. For a more extensive description of hypopigmenting agents and their modes of action, readers are referred to recent reviews on this topic [90,91].

3. Biological effectors

One mechanism to induce hypopigmentation in the skin is to inhibit crucial factors such as tyrosinase and other melanogenic enzymes, for example by targeting MITF [92], a key regulator of melanocyte function. In this context, a number of biological compounds, including transforming growth factor (TGF-β1) [93], tumor necrosis factor (TNFα) interleukins 1 and 6 (IL-1, IL6) [94], dickkopf 1 (DKK1) [4], calpain inhibitors [95], lysophosphatidic acid [96] and C2 ceramides, are able to inhibit the function of tyrosinase and related enzymes (TYRP1 and DCT), mainly through down-regulation of MITF.
Hypopigmenting properties of unsaturated fatty acids (e.g. linoleic acid) result from increased ubiquitination of tyrosinase that decreases its enzymatic function [97].

Another target is MC1R, as loss-of-function mutations in this receptor result in red hair and fair skin. As already mentioned, the physiological antagonist ASP blocks the binding of αMSH to MC1R and initiates a switch from eu- to pheo-melanogenesis. However, the specific mechanism regulating this switch has yet to be elucidated. The downstream target of MC1R, e.g. cAMP levels, can be decreased by androgens in combination with sex-hormone binding globulin, although drawbacks to this approach include a weak hypopigmenting effect and side effects of the agents.

Another possible way to inhibit skin pigmentation is by decreasing the transfer of melanosomes to keratinocytes. As mentioned above, the PAR2 receptor on keratinocytes plays an important role in regulating the uptake of melanosomes by keratinocytes. RWJ-50353, a serine protease inhibitor, leads to depigmentation in reconstructed skin and in dark-skinned Yucatan swine by affecting melanosome transfer and distribution [98]. In vitro, centaureidin [99], a flavone from Achillea millefolium (common yarrow), and methylphiophogonanone B [100] (MOPB), a homoisoflavonoid from Ophiopogon japonicus (mondo grass), block melanosome transfer by inducing the retraction of melanocyte dendrites through activation of GTPase Rho (the master regulator of dendrite formation in melanocytes) [101]).

Niacinamide (the amide form of vitamin B3), inhibits melanosome transfer in vitro and has a significant effect in reducing hyperpigmentation in vivo, although its mode of action is not known [102]. Another in vitro study showed that plasma membrane lectins and their glycoconjugates interfere with melanocyte-keratinocyte interactions by binding to their specific plasma membrane receptors and inhibiting melanosome transfer [103].

Inducing melanocyte destruction is another approach to induce skin hypopigmentation. Tryptophan metabolites from malassezia yeast induce apoptosis in human melanocytes, which leads to depigmentation of the skin (pityriasis versicolor) [104]. Imiquimod, an immune response modifier that stimulates cytotoxic T-cell-mediated responses through activation of toll-like receptors, leads to tumor destruction and depigmentation in lentigo-maligna lesions [105,106].

Imatinib mesylate, a drug used to treat chronic myeloid leukemia, has been reported to induce vitiligo-like depigmentation of the skin and it was suggested that the impairment of pigmentation involved SCF and its receptor c-kit [107,108].

4. Chemical effectors

For many years, the complex mechanisms involved in the abnormal up-regulation of melanocyte function were not well understood. Therefore, therapeutic chemical hypopigmenting agents are restricted to mainly inhibitory or cytotoxic effects. Since good progress has been made in elucidating the paracrine and autocrine networks involved in pigmentary disorders, further development of new selective agonists and antagonists for specific targets is anticipated.

Many of the chemical agents used to induce hypopigmentation affect tyrosinase function. Phenolic compounds are widely used, such as hydroquinone (HQ), which is considered one of the most effective
inhibitors of melanogenesis and is regarded as a reference standard when depigmenting agents are evaluated. HQ decreases tyrosinase activity by ~90% via the generation of quinones and reactive oxygen species (ROS) that induce damage in membrane lipids and proteins, by interacting with copper at the active site of the enzyme and by affecting RNA and DNA synthesis as well melanosome function [109]. In spite of its relatively high melanocyte-specific toxicity that improves hyperpigmentation in 14–70% of cases [110] and its general safety, there are common side effects of HQ such as skin irritation, contact dermatitis and (rarely) ochronosis, a blackish hyperpigmentation very resistant to treatment. The related phenol HQ monobenzyl ether (MBEH), which is used to eliminate residual spots of normal pigmentation in patients with generalized vitiligo, is metabolized to free radicals intracellularly which leads to selective melanocyte destruction and competitive tyrosinase inhibition, and results in an irreversible depigmentation even at sites that are distant from the application site.

Another common agent, arbutin, a natural β-glycoside of HQ isolated from the fruit of the California buckeye (Aesculus californica), as well as its more potent synthetic α-glycoside form, and synthetic deoxyarbutin are very efficient tyrosinase inhibitors. Compared to HQ, deoxyarbutin induces a more prolonged depigmentation and is not such an irritant.

Kojic acid, a fungal metabolic product from Aspergillus and Penicillium species, is commonly used in Asia as a whitening agent and a diet supplement, and it acts as a tyrosinase inhibitor by chelating copper at the active site of the enzyme [111]. Although it is effective as a hypopigmenting agent [112], especially in combination with other substances, it has a high sensitizing potential and can cause contact dermatitis [113]. Concerns have been raised about a possible carcinogenic effect since kojic acid was associated with hepatic tumors when fed to p53 deficient mice [114]. Aloesin, a natural derivative of aloe vera, acts as a competitive inhibitor of DOPA oxidation and a non-competitive inhibitor of tyrosine hydroxylase activity [115]. In a clinical trial, a combination of aloesin and arbutin inhibited UV-induced melanogenesis [116].

Azelaic acid, a dicarboxylic acid isolated from cultures of pityrosporum ovale, exerts antiproliferative and cytotoxic effects by inhibiting mitochondrial oxidoreductase and DNA synthesis [117] in highly active (or abnormal) melanocytes and acts as a weak inhibitor of tyrosinase activity. Its efficacy in treating hypermelanosis, which has been reported to be as good or even better than HQ [118], has been confirmed by several clinical studies [119].

Loss of melanin in the epidermis can also be obtained by stimulating the desquamation of stratum corneum cells. Retinoic acids are the most important agents in this context and act by disrupting pigment transfer, by inhibiting the dispersion of pigment granules in keratinocytes, by accelerating epidermal turnover and inducing desquamation [120]. Topical tretinoin has efficacy in treating hyperpigmentary disorders, although side effects such as erythema, peeling and PIH have been reported. To increase efficiency, various exfoliants (e.g. pretreatment with topical tretinoin followed by peels with trichloroacetic acid or α-hydroxy acids) can be combined. Another approach to increase efficacy is to combine different hypopigmenting agents that interfere with distinct steps of the melanogenic pathway (e.g. HQ and tretinoin).
Challenges for the future

As mentioned above, phenolic compounds are widely used as depigmenting agents, but it has been reported that instead of skin lightening, undesired counter-effects can occur. The flavone quercetin, originally described as a tyrosinase inhibitor [121], turned out to be a strong inducer of melanogenesis in vitro [122]. Although some hydroxylated derivatives of coumarins were reported to inhibit melanogenesis [123], several coumarins of seven umberiferae plants stimulated melanogenesis in murine melanoma cells [124]. The number of putative hypo- or hyper-pigmenting agents is vast, but promising effects detected in vitro are often not confirmed in clinical tests. Although topical application of compounds is most preferable, there are limitations to transdermal delivery as compounds applied to the surface of the skin are 100–1000 times diluted by the time they reach the lower layers of the epidermis [125]. Systemically applied substances carry the risk of greater side effects on other tissues. For pigmentation enhancing drugs, only MSH analogs have been tested extensively on humans, but there are concerns about their safety as injections with these synthetic compounds have been associated with nausea, facial flushing, fatigue and spontaneous erections [78,126].

Some critics have voiced their doubts whether enhancing pigmentation will significantly increase protection from skin cancers, as tanned skin has a sun protection factor (SPF) of only 2–4 and other physiological features might play a more important role in photoprotection. In addition, despite the fact that endogenous pigmentation is associated with a reduced cancer risk, it has been difficult to prove that facultative pigmentation has the same effect. Finally, there are some major concerns about the safety of artificially triggering the tanning response via the MC1R pathway since the stimulation of melanogenic activity also increases proliferation and might have undesired carcinogenic effects.

Conclusions

Facing the continuous rise in rates of skin cancer among the fair-skinned population and given the fact that in terms of skin cancer, highly pigmented skin is up to 70-fold more protected against the deleterious effects of UVR than is fair skin, artificially increasing pigmentation in the skin is more than ever a spotlight topic.

Consumer-driven demand for depigmenting drugs has risen in the past years due to the aging population’s interest in treating lesions resulting from photo-aging and due to demographic changes that have led to increases in non-Caucasian populations that are generally more prone to pigmentary disorders.

A better understanding of the complex interactions between regulators of the melanogenic pathway and the development of novel approaches to modulate pigmentation followed by strictly controlled clinical trials to assess safety and efficacy of these drugs would be desirable.
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Nonstandard Abbreviations Used

ACTH  adrenocorticotropic hormone

cAMP  cyclic adenosine monophosphate

ASP  agouti signal protein

bFGF  basic fibroblast growth factor

DCT  DOPAchrome tautomerase

DHI  5,6-dihydroxyindole

DHICA  DHI-2-carboxylic acid

DKK1  dickkopf 1

DOPA  β-3,4-dihydroxyphenylalanine

ET  endothelin

HGF  hepatocyte growth factor

HQ  hydroquinone

IL  interleukin

LS  lentigines solares

MC1R  melanocortin 1 receptor

MITF  microphthalmia-associated transcription factor

αMSH  α-melanocyte stimulating hormone

NGF  nerve growth factor

OA1  ocular albinism type 1

OCA  oculocutaneous albinism

PG  prostaglandin
Footnotes

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